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Using an Olympus SZX12 stereoscopic microscope with a CCD camera or, in some cases, an Olympus IX71 inverted microscope with relief contrast objectives, the movement of the injected beads was observed in real-time and recorded with a VHS video recorder (Panasonic, Osaka, Japan) or a DP21 CCD camera controller (Olympus). Image-J software and its plug-in Color Footprint Rainbow were used to visualise bead movements as multi-coloured video-tracking images. Seeding and culture of U251 and U87MG stem cell-derived cells (SCs) a The U251 and U87MG cells were digested by 0.05% Trypsin and transferred to 6-well plates (1005 cells/well), and then cultured in DMEM/F12 medium supplemented with 10% FBS and 5g/ml BSA. After that, the cells were treated with 5M PNA-A15 or scramble for 48 h. The expressions of the stem cell markers NANOG and CD133 were determined by quantitative real-time PCR (RT-qPCR). b The SCs were dissociated into single cells by 0.05% Trypsin and then seeded into a 24-well ultralow attachment plate at a density of 1.05 to 2.05 cells/well. After incubation for 7 days, the images of the SC-spheroid formation were taken and evaluated using an inverted microscope (Olympus IX51 with DP70). The size of each SC-spheroid was calculated and their relative numbers were determined. All plots present the meanSD (n=3). *P

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Using a Nanoject instrument (Drummond Scientific, Broomall, PA, USA), Qdot565 solution containing 2 M quantum dot nanocrystals (Qtracker565 non-targeted quantum dots 2 M solution, catalog Q21031MP, Quantum Dot Co., Hayward, CA, USA) was injected. In the stock solution, Qdot565 was suspended in 50 mM borate buffer (pH 8.3), and the stock solution was injected without dilution. Injection volumes were 4.6 or 9.2 nL per embryo at stage 30 to 38, and 23 or 46 nL per larva at later developmental stages (stage 41-51), or for tadpoles with well-developed hind limbs (stage 53). Numbers injected were: stage 30-38, n = 9; stage 41-43, n = 19; stage 46-48, n = 19; stage 50-51, n = 12; stage 53, n = 8. The Nanoject specifications enable the injection of volumes with a precision of two significant digits. The injected larvae were observed using a fluorescence stereoscopic microscope (Olympus, SZX12, Tokyo, Japan), using excitation at 460 to 490 nm, and photographed with a digital camera (Olympus, DP-70). The expression of down- and upregulated genes containing A and T repeats (X-axis) with the repeat density in bp/Mbp (Y-axis). a Lysates of parental cells and CSC-enriched cells were prepared and analysed for CD133, CD44, ABCG2 and ALDH1A1 expression by Western blotting. The GSE142616 microarray dataset was downloaded from GEO. Only perfect A and T repeats were classified as downregulated (Dn), neutral (Nu), or upregulated (Up) genes. Experimental and control samples were the spheroid and monolayer of b H292 and c A549 cell lines, respectively. d Venn diagram shows overlap between the intersection of downregulated genes between H292 (the left circle) and A549 (the right circle) cell lines. e The list of candidate genes determined by biological process from Gene Ontology. Student's t-test P-values, which are denoted by P, indicate that the mean difference between Dn/Up and Nu is statistically significant. All plots present the meanSD (n=3). *P 5ec8ef588b

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